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Microbe profile

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Kraft, Beate; Canfield, Donald E.

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7 Microbe Profile: *Nitrosopumilus maritimus*

8 **Authors:** Beate Kraft^{1*} and Donald E. Canfield^{1,2,3,†}

9 **Affiliations:**

10 ¹Nordcee, Department of Biology, University of Southern Denmark, Odense, Denmark

11 ²Key Laboratory of Petroleum Geochemistry, Research Institute of Petroleum Exploration and

12 Development, China National Petroleum Corporation, Beijing 100083, China

13 ³Danish Institute of Advanced Study, 5230 Odense, Denmark

14 *corresponding author: bkraft@biology.sdu.dk

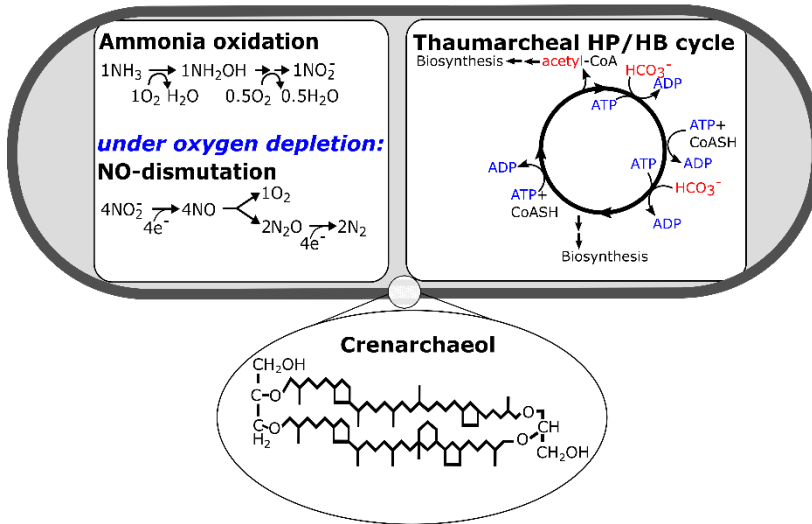
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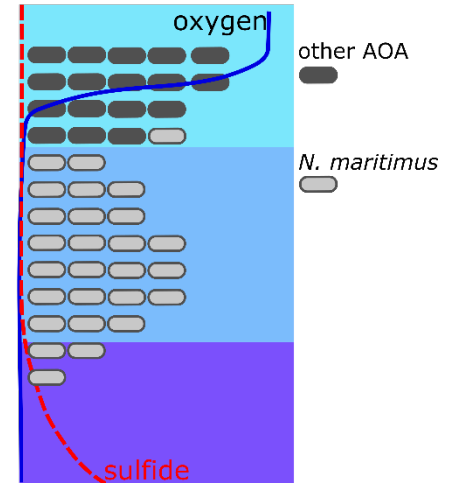
17 **Graphical abstract**

Nitrosopumilus maritimus: A marine ammonia-oxidizing archaeon

A) Properties and Physiology



B) Ecology



18

19 **A)** *Nitrosopumilus maritimus* gains energy by ammonia oxidation coupled to oxygen consumption. When
 20 oxygen is depleted, it produces its own oxygen. NO-dismutation is the proposed oxygen production pathway.
 21 Carbon fixation operates via a modified hydroxypropionate/hydroxybutyrate cycle. Crenarchaeol is a
 22 membrane lipid unique to ammonia-oxidizing archaea (AOA). **B)** Distribution of *N. maritimus*-related AOA in
 23 relation to oxygen concentrations in the example of the Black Sea [1].

24 **Abstract**

25 *Nitrosopumilus maritimus* is a marine ammonia-oxidizing archaeon with a high affinity for ammonia. It fixes
 26 carbon via a modified hydroxypropionate/hydroxybutyrate cycle and shows weak utilization of cyanate as
 27 supplementary energy and nitrogen source. When oxygen is depleted, *N. maritimus* produces its own oxygen,
 28 which may explain its regular occurrence in anoxic waters. Several enzymes of the ammonia oxidation and
 29 oxygen production pathways remain to be identified.

30 **Taxonomy:**

31 Domain *Archaea*, phylum *Nitrososphaerota* (Thaumarchaeota), class *Nitrososphaeria*, order
 32 *Nitrosopumilales*, family *Nitropumilaceae*, genus *Nitrosopumilus*, species *Nitrosopumilus maritimus*.

33

34 **Properties:**

35 *Nitrosopumilus maritimus* SCM1, isolated from a tropical marine fish tank at the Seattle Aquarium, is a
36 chemolithoautotroph, gaining energy from aerobic ammonia oxidation to nitrite [2]. The cells are non-motile
37 straight rods with a diameter of 0.17–0.22µm and a length of 0.5–0.9µm. The core membrane lipids of *N.*
38 *maritimus* consist of glycerol dialkyl glycerol tetraethers (GDGTs), including crenarchaeol. Since crenarchaeol
39 is unique to Nitrososphaerota, it is used as biomarker for this archaeal phylum including *Nitrosopumilus* spp..
40 *N. maritimus* produces methylphosphonate esters that can release methane when degraded, potentially
41 contributing to methane production in marine oxic waters [3].

42

43 **Phylogeny:**

44 Ammonia oxidizing archaea (AOA) form the monophyletic class *Nitrososphaeria*, which divides into the group
45 of *Candidatus* Nitrosocaldales and a second group that splits into two major lineages: the order
46 *Nitrososphaerales*, with representatives mostly in soils, and a second major clade. This clade consists of the
47 two orders *Candidatus* Nitrosotaleales, and *Nitrosopumilales*. The *Nitrosopumilales*, including the species
48 *Nitrosopumilus maritimus*, are mostly found in marine environments.

49

50 **Ecology**

51 Members of the order *Nitrosopumilales* are ubiquitous and abundant in the environment. They are key
52 players in the nitrogen cycle of the world's oceans and make up approximately 20% of the microbial
53 community in the oxic water column. They are abundant in oxygenated environments, but also found in
54 oxygen-depleted environments such as marine oxygen-minimum zones and anoxic basins like t the Black sea,
55 even though ammonia-oxidation requires oxygen [1].

56

57 **Key features and discoveries:**

58 **1. Ammonia oxidation**

59 *N. maritimus* SCM1 has an extremely high specific affinity for ammonia (defined here as ammonia +
60 ammonium) with an apparent half-saturation constant (*K_m*) of 0.132 µM [4]. A high ammonia affinity allows
61 members of the genus *Nitrosopumilius* to thrive in the open ocean, for example, where ammonium is present
62 at low nanomolar concentrations.

63 The biochemical pathway of ammonia oxidation in *N. maritimus* or other AOA is not fully resolved. With
64 current understanding, the first step of ammonia oxidation by *N. maritimus* is the oxidation of ammonia to

65 hydroxylamine, catalyzed by a putative ammonia mono oxygenase (AMO) [5]. The gene encoding for the
66 alpha subunit of AMO, *amoA*, is typically used to assess the diversity and abundance of AOA in the
67 environment. The enzyme responsible for hydroxylamine oxidation is unclear and genes encoding for a
68 homologue to the bacterial hydroxylamine dehydrogenase (HAO) are absent in the *Nitrososphaeria* including
69 *N. maritimus*. The *N. maritimus* genome, however, encodes several blue copper-containing plastocyanin-like
70 electron carriers. Some of them have been proposed to be involved in electron transfer from hydroxylamine
71 or other potential intermediates to the terminal oxidase [5].

72 **2. Utilization of alternative nitrogen substrates**

73 Marine *Nitrosospumilales* often live with vanishingly low ammonium concentrations and they can
74 supplement their nitrogen requirements with simple organic nitrogen compounds that are ubiquitous in
75 marine systems [6]. Pure cultures of *N. maritimus* SCM1 can convert cyanate to ammonia, using it as energy
76 and nitrogen source even though no known cyanases are present in its genome nor any other marine
77 *Nitrosopumilales*. While some *Nitrosopumilales* can use urea, *N. maritimus* SCM1 cannot, and its genome
78 does not encode any known ureases [6].

79

80 **3. Carbon fixation pathway**

81 *N. maritimus* assimilates inorganic carbon via a modified hydroxypropionate/hydroxybutyrate (HP/HB) cycle
82 distinct from the cycle operating in Crenarchaeota [7]. In the HP/HB cycle of *Nitrososphaeria*, ADP (and not
83 AMP as in the Crenarchaeota) is produced during the activation of 3-hydroxypropionate and 4-
84 hydroxybutyrate. Furthermore, some enzymes catalyze multiple reactions, which reduces the cost of protein
85 biosynthesis. Therefore, the HP/HB cycle in *Nitrososphaeria* is the most energy efficient carbon fixation
86 pathway found in aerobes, helping *Nitrososphaeria* to attain high numbers in oligotrophic environments of
87 low energy supply and ammonia limitation.

88 While the growth of *N. maritimus* is stimulated by the small organic molecules pyruvate, oxaloacetate, and
89 α -ketoglutarate, mixotrophy has not been confirmed [8]. α -Keto acids abiotically scavenge hydrogen
90 peroxide. *N. maritimus* lacks the hydrogen peroxide-detoxifying enzyme catalase [9], and by removing
91 hydrogen peroxide, α -Keto acids may enhance growth.

92 **4. NO-dismutation and oxygen production**

93 *N. maritimus* SCM1 has a relative low affinity for oxygen with a relatively high apparent half saturation
94 constant of $K_m = 3.9 \mu\text{M}$ [4]. With such a high K_m , *N. maritimus* should have difficulties competing with other
95 aerobes utilizing high-affinity oxidases in oxygen-limited environments. We recently showed, however, that

96 *N. maritimus* SCM1 can produce its own oxygen when exposed to anoxia [10], and the oxygen produced is
97 partly used for ammonia oxidation. In the proposed oxygen-production pathway (see graphical abstract), *N.*
98 *maritimus* reduces nitrite to NO via a NirK-nitrite reductase. We proposed that NO is then dismutated to
99 oxygen and nitrous oxide, which is further reduced to N₂. Producing one oxygen molecule requires four nitrite
100 molecules, and the coupling of NO-dismutation to ammonia oxidation would lead to a net loss of nitrite. The
101 proposed NO-dismutation pathway in *N. maritimus* would constitute the only known oxygen production
102 pathway in the archaeal domain. The proposed pathway in *N. maritimus* is similar to the NO-dismutation
103 pathway proposed for the methanotroph *Candidatus Methylopirabilis oxyfera* [11] in that each organism
104 produces oxygen for the aerobic oxidation of their key metabolic electron donor (ammonia or methane).
105 However, intermediates of the pathways seem to differ, and oxygen accumulates in the medium of *N.*
106 *maritimus*, with the potential to support other aerobes in the environment.

107 **Open questions:**

- 108 • Several enzymes catalyzing steps in the ammonia-oxidation and NO-dismutation pathways remain
109 unidentified. The identification of these missing enzymes is a crucial for elucidating the pathways and
110 their intermediates.
- 111 • So far, oxygen production has only been shown in pure-culture incubations of *N. maritimus*. Is the dark
112 oxygen production pathway present in other AOA? What is the ecological relevance of NO-dismutation
113 and oxygen production by AOA?
- 114 • What are the interactions of *N. maritimus* with other microbes in the environment? For example, can the
115 oxygen released by *N. maritimus* support other aerobes in anoxic environments?

116
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120

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